



CaptuVir100

DEVICE INFORMATION



1. DESCRIPTION

CaptuVir100 is a ready-to-use device for the purification of large biopharmaceuticals such as viruses, virus vectors, and virus-like particles (VLPs) by membrane-based steric exclusion chromatography (SXC).

SXC technology offers size-based selectivity to concentrate, purify, and buffer-exchange a wide variety of viruses relevant in gene therapy and vaccine applications.

Benefits of membrane-based SXC:

- **Higher product yields:** 20–50% higher product yield compared to other chromatography methods and resins mean less wasted product and smaller columns needed for purification
- **One-size-fits-all recipe:** a single recipe for a wide variety of viruses and serotypes shortens process development & deployment
- **Faster processes:** flow rates 10–20× higher compared to resins thanks to convective flow
- **Single-use:** avoids impurity carry-over and cross-contamination and reduces CapEx & OpEx
- **Increased product stability:** avoids extreme pH and salt concentrations
- **Scalability:** increases capacity by increasing membrane surface

For more information, see the QR code or visit <https://www.contivir.com/dsp-sxc>



2. TECHNICAL INFORMATION

Stationary phase	Cellulose membranes
Total surface	100 cm ²
Device volume (DV)	1.0 mL
Minimal capacities per device	$\approx 3 \times 10^{12}$ vg ^a (AAV) $\approx 6 \times 10^9$ PFU ^b (yellow fever virus) $\approx 3.7 \times 10^9$ TCID ₅₀ ^c (MVA virus) $\approx 1.3 \times 10^6$ TCID ₅₀ ^c (VSV-NDV) $\approx 9 \times 10^5$ HAU ^d (influenza virus) $\approx 47 \mu\text{g}_{\text{HA}}$ ^e (influenza virus) $\approx 4 \times 10^{10}$ IU ^f (hepatitis C virus)
Recommended flow rate	5 mL/min (max. 10 mL/min)
Maximum operating pressure	2.0 MPa (20 bar)
pH stability	2–12 (operational) 2–14 (cleaning-in-place)
Storage	20% (v/v) ethanol in water 2–8°C

a vg: viral genomes; AAV: adeno-associated virus

b PFU: plaque forming units

c TCID₅₀: tissue culture 50% infective dose; MVA: Modified Vaccinia Ankara; VSV: vesicular stomatitis virus; NDV: Newcastle disease virus

d HAU: hemagglutination activity units

e HA: hemagglutinin antigen

f IU: international units

3. HOW TO USE

CaptuVir100 can be used manually, with a peristaltic pump or a chromatography system. Operation with a chromatography system is highly recommended.

Reagents

- **Binding buffer:** 16% PEG-6000, PBS 1× (for viruses of >80 nm) or 20% PEG-6000, PBS 1× for viruses of <80 nm)
- **Equilibration/Wash buffer:** dilute Binding buffer 1:1 with PBS 1×
- **Elution buffer:** PBS 1×

Device and method set-up

- Connect the device to the appropriate system ports using fingertight 1/16" male connectors and PEEK tubing of 1 mm internal diameter.
- Program a new method in the chromatography system. If choosing from a pre-defined method, an affinity purification template with a step elution is appropriate.

Experimental procedure

- **Clarify the crude virus solution** by centrifugation and microfiltration.
- **Equilibration:** Flush the device with ≥ 10 DV of Equilibration/Wash buffer.
- **Sample loading:** a 1:1 in-line dilution of the clarified virus harvest with the Binding buffer is preferable over conditioning the sample with PEG off-line
- **Wash:** flush the device with ≥ 10 DV of Equilibration/Wash buffer or until UV signal is baseline and stabled.
- **Elution:** recover the product by flushing the device with PBS 1×

For more information, see the QR code or visit
<https://www.contivir.com/captuvir100>

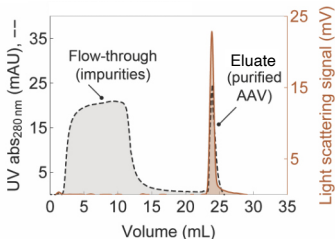


4. EXAMPLE

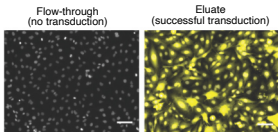
CaptuVir100 can be used to purify different AAV particles with >95% product yield from both cell lysates and supernatants using the same process conditions. At least 10 different AAV serotypes and display mutants have been purified, including: AAV1, AAV2, AAV6, AAV8, AAV9, and AAV-DJP2.

The diagram below shows a chromatogram (A), a transduction assay of the purified eluate (B) and a TEM picture (C) of the purified AAV particles and illustrates the successful purification of biologically active AAV.

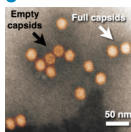
A



B Cell transduction from SXC fractions



C TEM picture





www.contivir.com

**Enabling Bedside
Manufacturing of Viral Vaccines
& Gene Therapies**

ContiVir is a research project of the Max Planck Institute for Dynamics of Complex Technical Systems.

Max Planck Institute for Dynamics of Complex
Technical Systems
Sandtorstrasse 1
39106 Magdeburg
Germany

This device is for research use only. It is not intended for use in any clinical or diagnostic procedures.

The Max-Planck-Institute Magdeburg, the Max-Planck Society or any of its employees are not liable for any direct or indirect losses or damages arising out of or in connection with the use of this device.

The user is responsible for the use of this device.